

# Finding Bermuda buttercup Shifts in reproductive traits after introduction in the Mediterranean basin

Sílvia Castro<sup>1</sup>, Peter Glasnović<sup>2</sup> et al.

<sup>1</sup>FLOWer Lab • Centre for Functional Ecology – Science for People & the Planet • Department of Life Sciences • University of Coimbra, Coimbra, Portugal

<sup>2</sup>Natural Sciences and Information Technologies • Faculty of Mathematics • University of Primorska, Koper, Slovenia













STUDY SYSTEM: Oxalis pes-caprae L.

# Geophyte

# **Asexual reproduction**

- High production of bulbs
- Root contraction capacities

# **Sexual reproduction**

Tristylous species

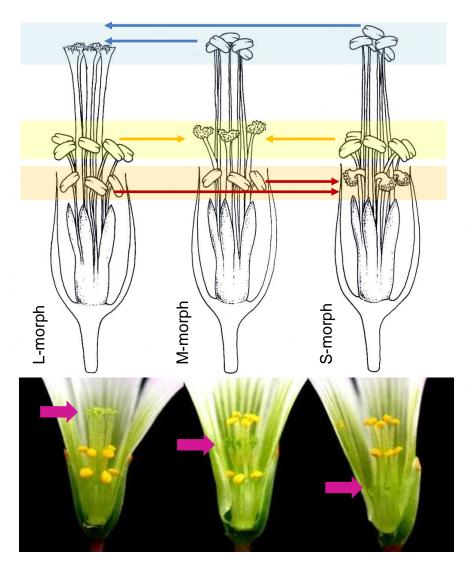
# **Polyploid complex**

• 2x, 4x and 5x cytotypes



# STUDY SYSTEM: Oxalis pes-caprae L.

#### **Sexual system**



# **Tristylous species**

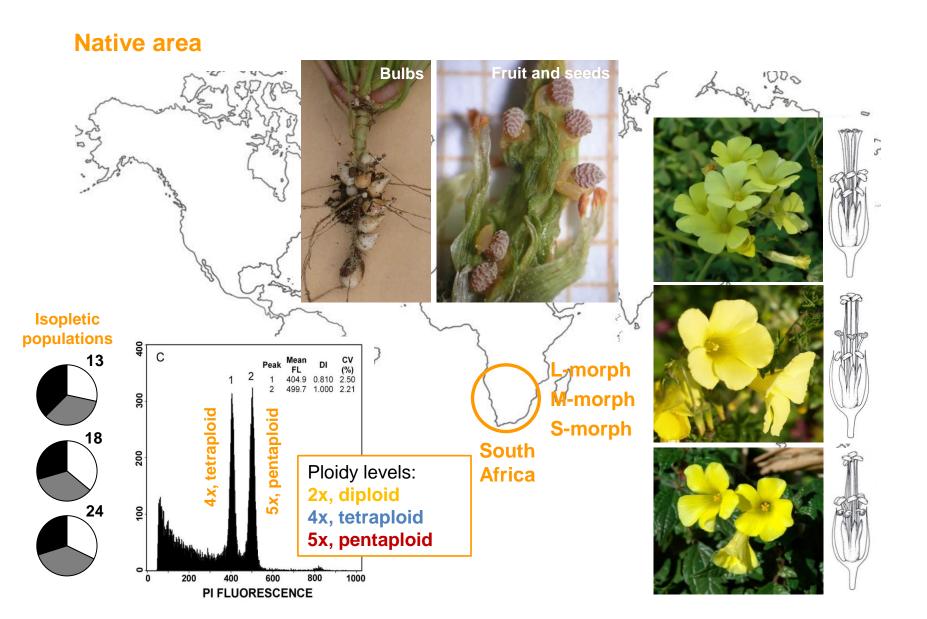
- Long-styled morph
- Mid-styled morph
- Short-styled morph

# **Trimorphic incompatibility**

(i.e., self and intra-morph incompatible)

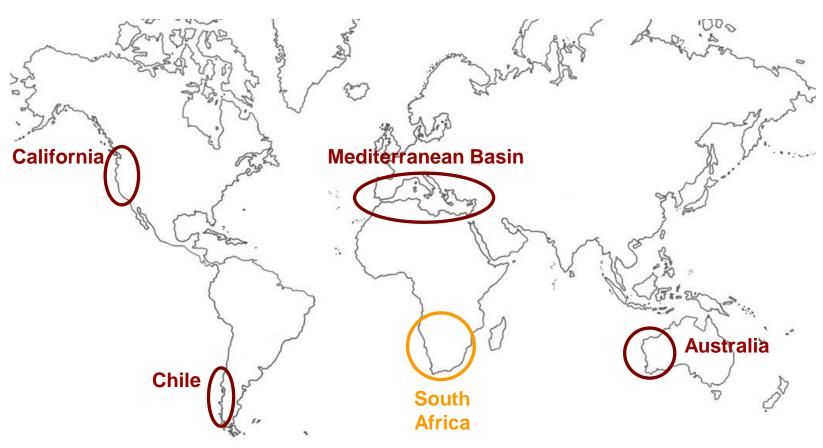
**Ornduff R. 1987.** Annals of Missouri Botanical Garden 74:79-84.

## Oxalis pes-caprae L.: DISTRIBUTION PATTERNS AND DIVERSITY OF FORMS



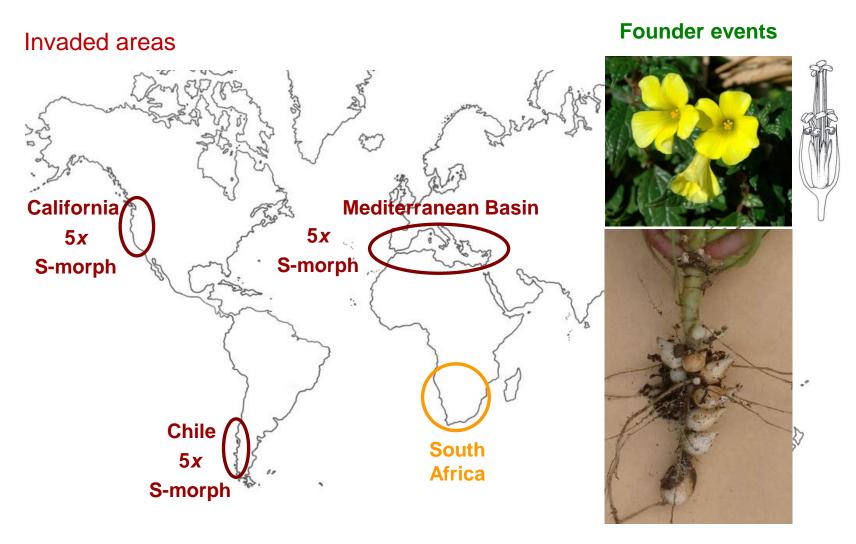
## Oxalis pes-caprae L.: DISTRIBUTION PATTERNS AND DIVERSITY OF FORMS

## Invaded areas



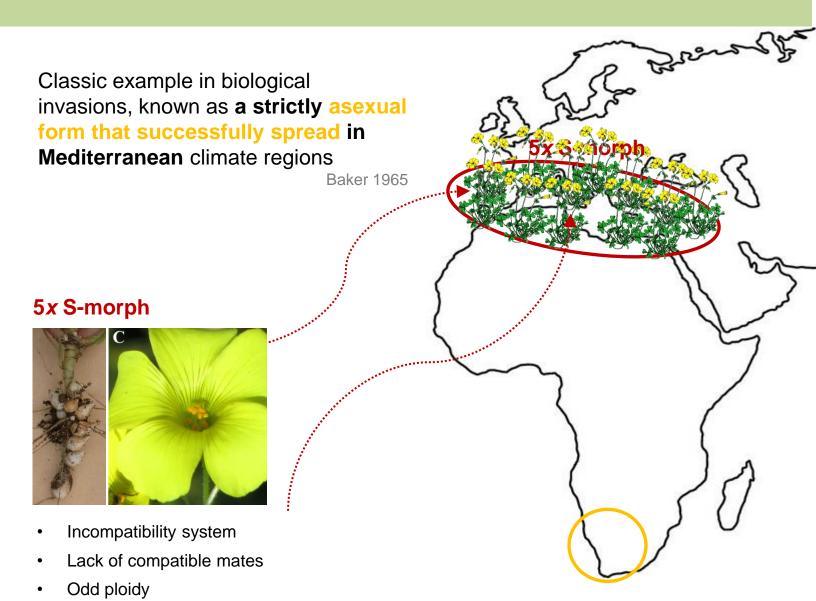
• Invasive success seems to be highly correlated with anthropogenic activities

#### Oxalis pes-caprae L.: DISTRIBUTION PATTERNS AND DIVERSITY OF FORMS

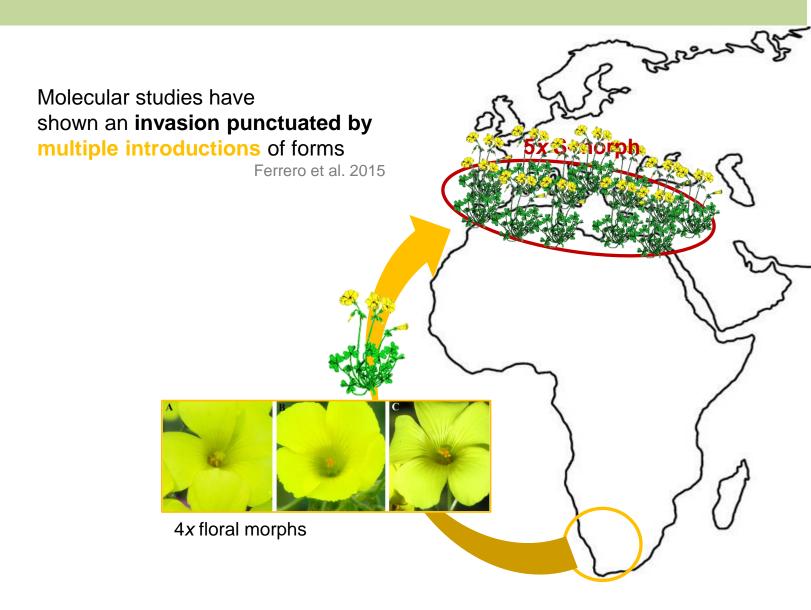


• Clonality clearly played a major role in invasion and enabled growth, persistence and spread after the absence of compatible mating partners.

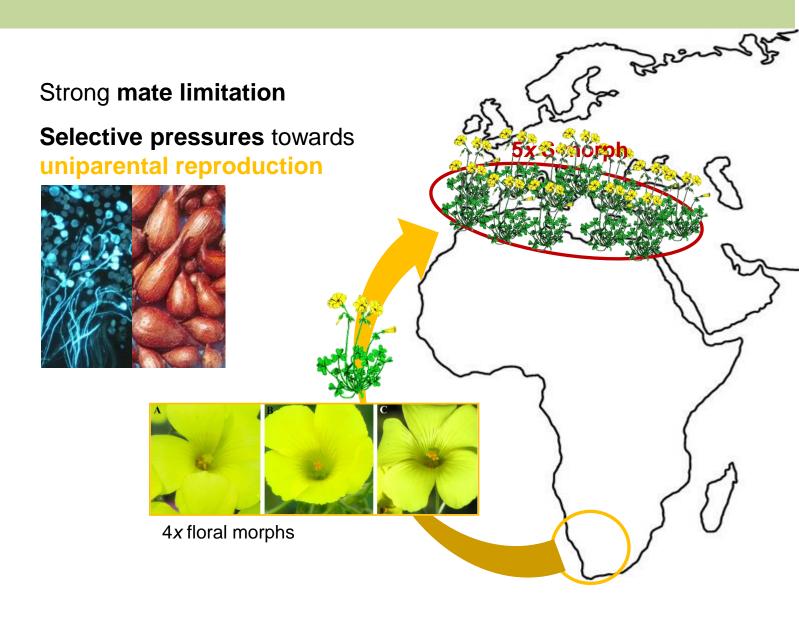




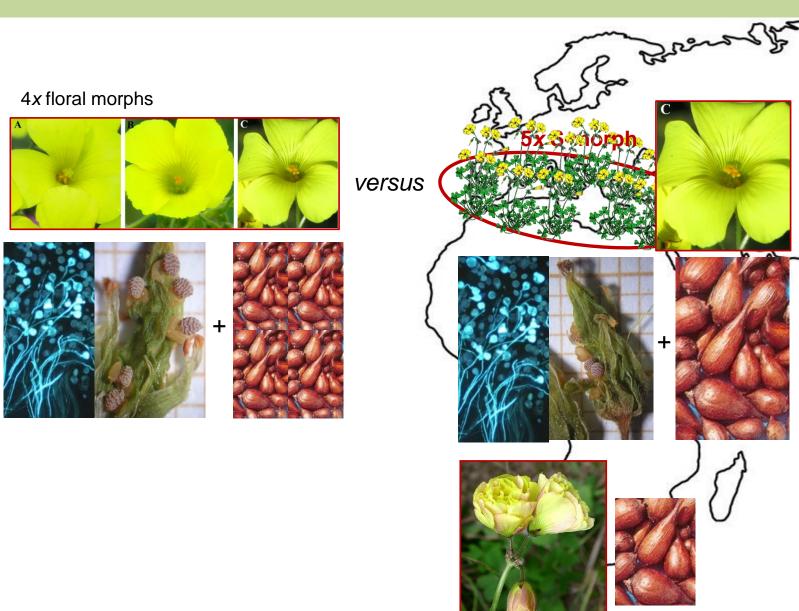




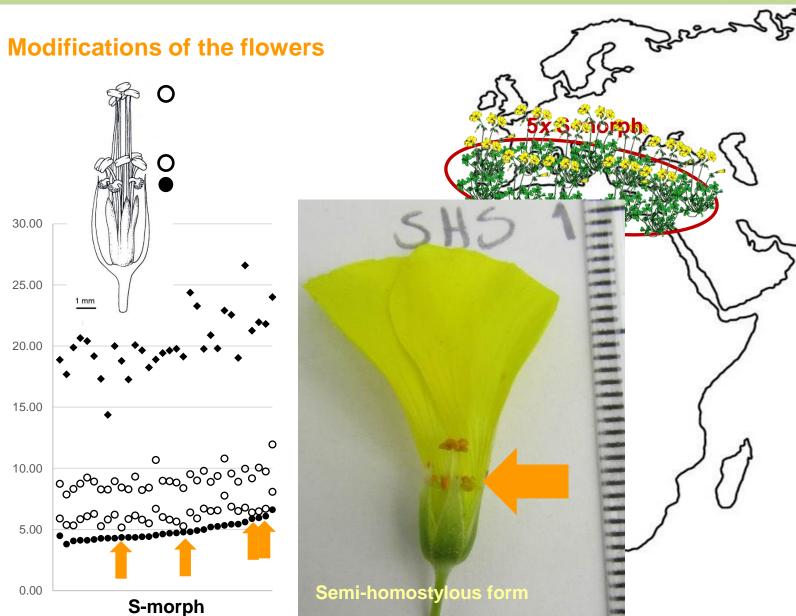














Bermuda buttercup invasion history: what next?

#### **AIM**

Understand the ubiquity of the changes in sexual strategies and assess the impact in the maintenance of flower polymorphisms by extending the sampling to other regions of the Mediterranean basin

Finding Bermuda buttercup





# Finding Bermuda buttercup

## **CALL FOR COLLABORATION**

You or any of your colleagues are **invited** to participate in this project



# Finding Bermuda buttercup

**PROTOCOL** 



#### 1. POPULATION CHARACTERIZATION

Population information Floral morph proportions Database



#### 2. PLANT SAMPLING FOR CYTOTYPE AND FITNESS

Morph voucher
Reproductive fitness samples
Cytotype samples
Genetic samples

#### 3. FLOWER MORPHOMETRY

Flower samples



#### 1. POPULATION CHARACTERIZATION

# **Population information**

- Geographical coordinates (WGS84)
- Habitat type (Table 1)
- Crop (if applicable)
- Soil movement
- Notes



#### Table 1.

Natural or semi-natural areas

Agricultural areas

- Intensive permanent crops
- Traditional permanent crops
- Annual crops

Managed forest areas

Managed urban areas

Non-managed urban areas

Managed peri-urban areas

Non-managed peri-urban areas

Rural areas

Road edges

Other

#### **Example**

- 39.577621, -9.054498
- Annual crops
- Crop: corn
- Yes
- Notes: plant remnants indicate a corn field

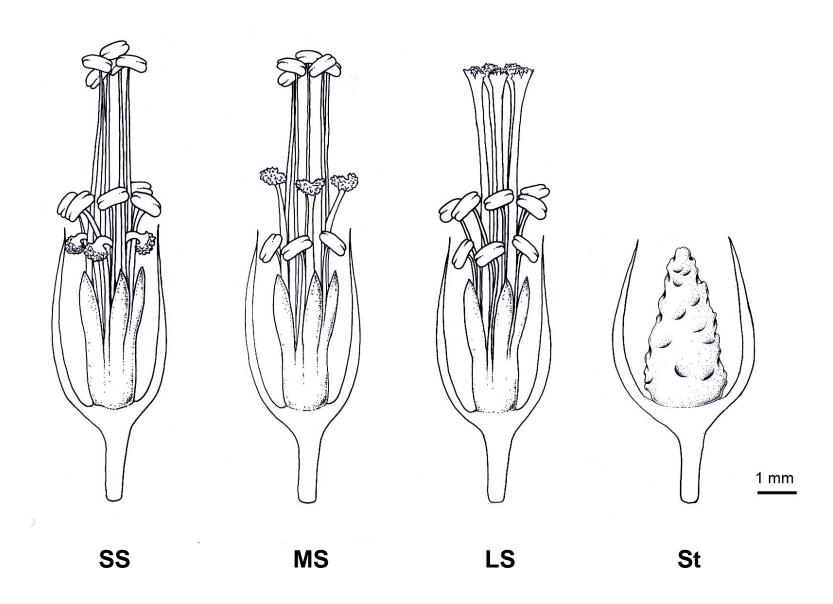


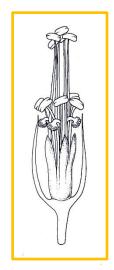
#### 1. POPULATION CHARACTERIZATION

# Floral morph proportions

 Observe the morph of at least 100 individuals (in 5-m intervals) across 2-3 longitudinal transects across the entire population





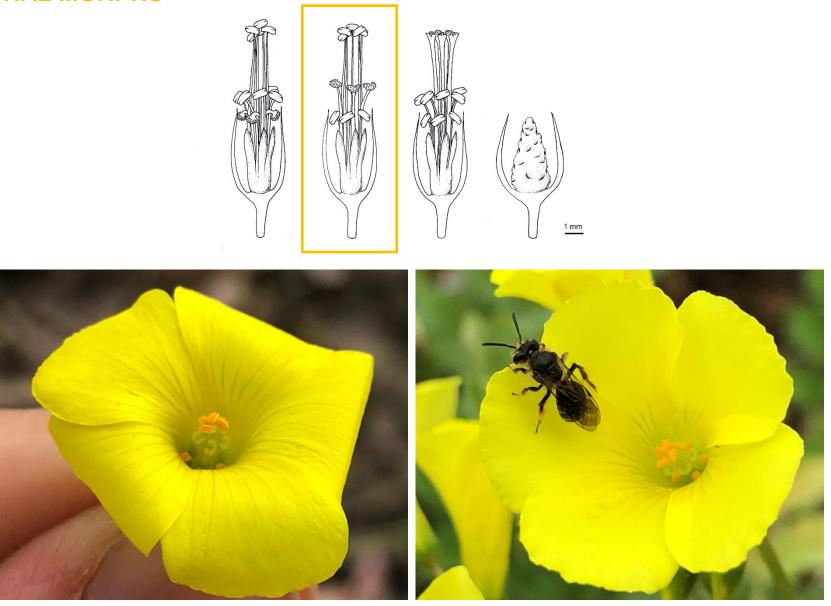




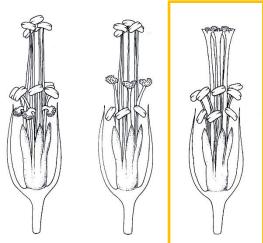




**Short-styled morph (SS)** 



Mid-styled morph (MS)

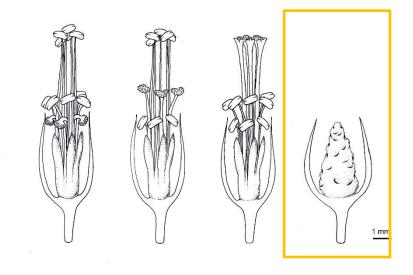






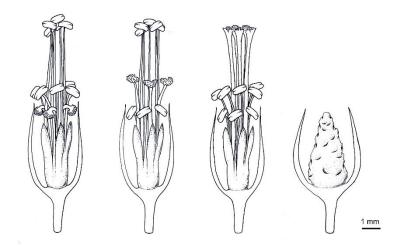


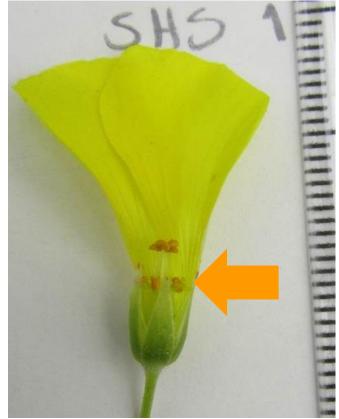






Sterile morph (St)





**Semi-homostylous morph (SHS)** 



# 1. POPULATION CHARACTERIZATION

**Database:** Enter the data in the Excel file provided

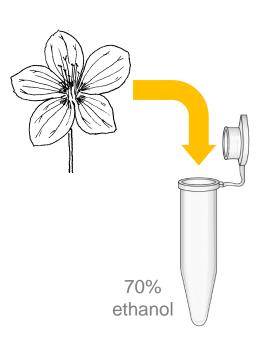
Α	В	С	D	Е	F
Excel Sheet - POPULATION Database				Excel Sheet - FITNESS Database	
COLUMN NAME	DESCRIPTION			COLUMN NAME	DESCRIPTION
ID	Identification code of the locality being sampled. Please use the following coding instructions: Country code_Researcher code_population number_plant number. e.g., PT-SC-25 refers to the population 25 sampled by Silvia Castro in Portugal.			ID	Identification code of the locality being sampled. Please use the following coding instructions: Country code_Researcher code_population number_plant number. e.g., SC-25 refers to the population 25 sampled by Sílvia Castro in Portugal.
Locality	Lowest administrative unit at the country level where the observations were undertaken			Locality	Lowest administrative unit at the country level where the observations were underta
Country	Country where the observations were undertaken			Country	Country where the observations were undertaken
Coordinates	Geographical coordinates provided in World Geodetic System 84 system			Plant ID	Number identifiyng the individual plant being sampled
Oxalis coverage (%)	Aproximate percentage of Oxalis pes-caprae coverage in the locality			Morph	Floral morph of the individual plant being sampled
Population area (m²)	Estimated area occupied by the population in square meters. If the population is a continuous in the region please indicate 'continuum' in column 'Notes'			No. flower buds	Number of flower buds in the selected flowering/fruting stem
Habitat	Characterize the habitat in the following categories: (Table 1 and Excel Sheet "Table 1 HABITAT"). Any additional information can be provided in the field "Notes"			No. flowers	Number of open flowers (including recently open flowers to wiltered flowers) in the selected flowering/fruting stem
Сгор	If the population is in agricultural area please indicate the crop			No. fruits	Number of developed fruits in the selected flowering/fruting stem
Soil movement	YES/NO and should refer to the occurence of soild movement during the last year			No. scars	Number of scars corresponding to the number of aborted flowers (i.e., flowers that on the developed into fruit) in the selected flowering/fruting stem
Co-flowering plants	List co-flowering plant species present at the locality				
Floral forms	Number of plants of each floral form observed in the transects across the population. Floral forms:				
	SS - Short-styled				
	MS - Mid-Styled				
	LS - Long-styled				
	SHS - Semi-homostylous				
	St - multipetalous sterile form				
Notes	Please use this space to add any note you may find pertinent				



#### 2. PLANT SAMPLING FOR CYTOTYPE AND FITNESS

Locate 10 plants from each morph and collect the following samples in every plant:

 Morph voucher: collect 1-2 flowers per plant to separate microtubes with ethanol 70% (1 microtube per plant) labelled inside with population and plant codes





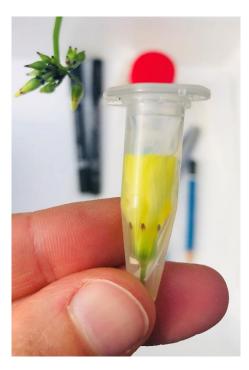




#### 2. PLANT SAMPLING FOR CYTOTYPE AND FITNESS

Locate 10 plants from each morph and collect the following samples in every plant:

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#### 2. PLANT SAMPLING FOR CYTOTYPE AND FITNESS

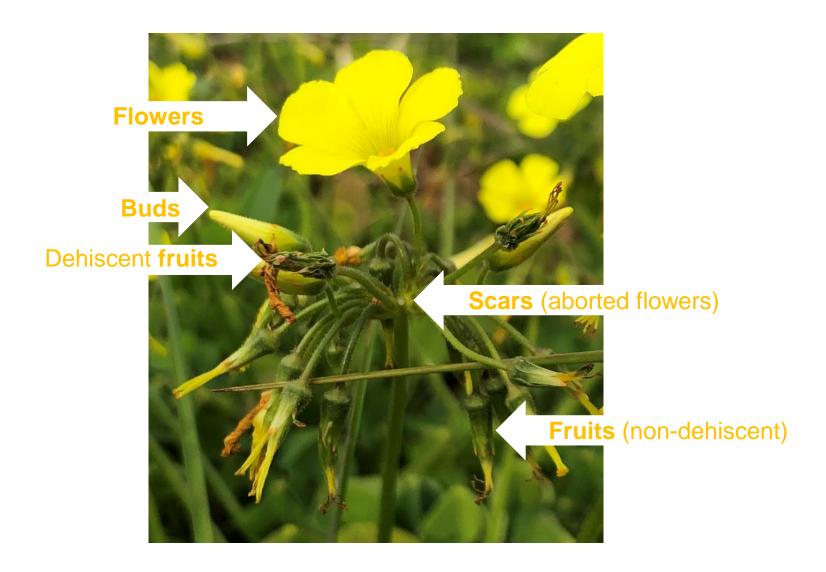
Locate 10 plants from each morph and collect the following samples in every plant:

- Reproductive fitness: you can proceed in one of two ways
- 1) identify a fruiting stem per plant and count the number of scars (i.e., aborted flowers), fruits, flowers and flower buds (if applicable); collect **3 non-dehiscent fruits** (when available) into a paper bag identified with the population and plant codes; **OR**
- 2) collect a fruiting stem per plant into a paper bag identified with the population and plant codes.

Notes: it is likely that fruits do not exist, thus, look for inflorescences with scars indicative of zero fruits

Place the bags to dry at 30 °C until completely dried.

# **SEXUAL REPRODUCTIVE FITNESS**



# **SEXUAL REPRODUCTIVE FITNESS**

# Fruits (non-dehiscent)



Fruits (after dried in paper envelopes)





#### 2. PLANT SAMPLING FOR CYTOTYPE AND FITNESS

Locate 10 plants per morph and collect the following samples in every plant:

 Cytotype analyses: collect the plant with some root system, prune the old leaves, store it in a small plastic bag identified with population and plant codes and stored at 4 °C

**IMPORTANT:** fresh samples need to be sent as fast as possible for flow cytometric analyses to:

#### Sílvia Castro

Department of Life Sciences, University of Coimbra, Calçada Martim de Freitas, 3000-456 Coimbra, Portugal

# **SAMPLES FOR FLOW CYTOMETRY**







**Root system** 

Label



#### 2. PLANT SAMPLING FOR CYTOTYPE AND FITNESS

Locate 10 plants per morph and collect the following samples in every plant:

 Genetic analyses: collect 4-6 young and healthy leaves per plant and store in separate paper envelopes with silica gel, labelled with population and plant codes

Notes: all the sampled plants should be separated at least 5-m apart to avoid re-sampling clones





#### 3. FLOWER MORPHOMETRY

- Collect at least 25 flowers from each morph to a vial with 70% ethanol labelled inside with population and plant codes
- Different morphs should be stored in separate vials



Send the flower samples to:

## Peter Glasnović

University of Primorska, Faculty of Mathematics, Natural Sciences and Information Technologies, University Campus Livade, Livade 4, 6310 Izola, Slovenia



#### **POPULATION CODING**

Country code\_Researcher code\_population number e.g., **PT-SC-25** refers to the population 25 sampled by Sílvia Castro in Portugal.

#### **PLANT CODING**

Country code\_Researcher code\_population number\_plant number e.g., **PT-SC-25-03** refers to the plant number 3 from the population 25 sampled by Sílvia Castro in Portugal.







NOTE: flowering period may be different in your region

#### TIMELINE

2022 Oct • WG1 Meeting to **present** the project

Nov • Development of the final field protocol

- Participants manifestation of interest (11 Nov)
- Dec Workshop for protocol demonstration and questions

2023 Jan • Sample collection (Dec 2022 to Mar 2023)

Fev • FCM analyses (Dec 2022 to Mar 2023)

Mar

Apr • Flower analyses in the lab (Jan-Jul 2023)

May

Jun • Preliminary data analyses (Jun-Jul 2023)

Jul

Ago • Presentation of results and next steps (MC Meeting)



Bermuda buttercup invasion history: what next?

#### **CALL FOR COLLABORATION**

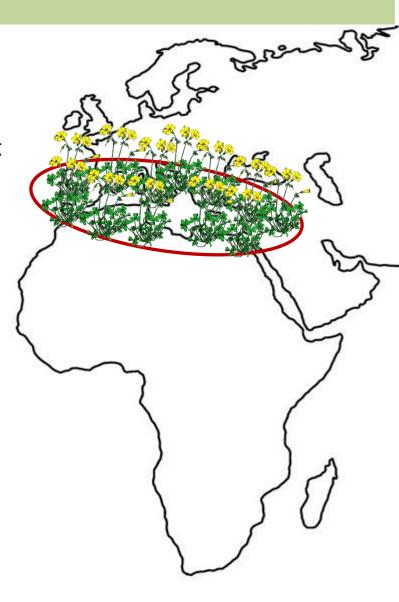
You or any of your colleagues are **invited to participate** in this project

MANIFESTATION OF INTEREST until 11th November

#### **OUTPUTS**

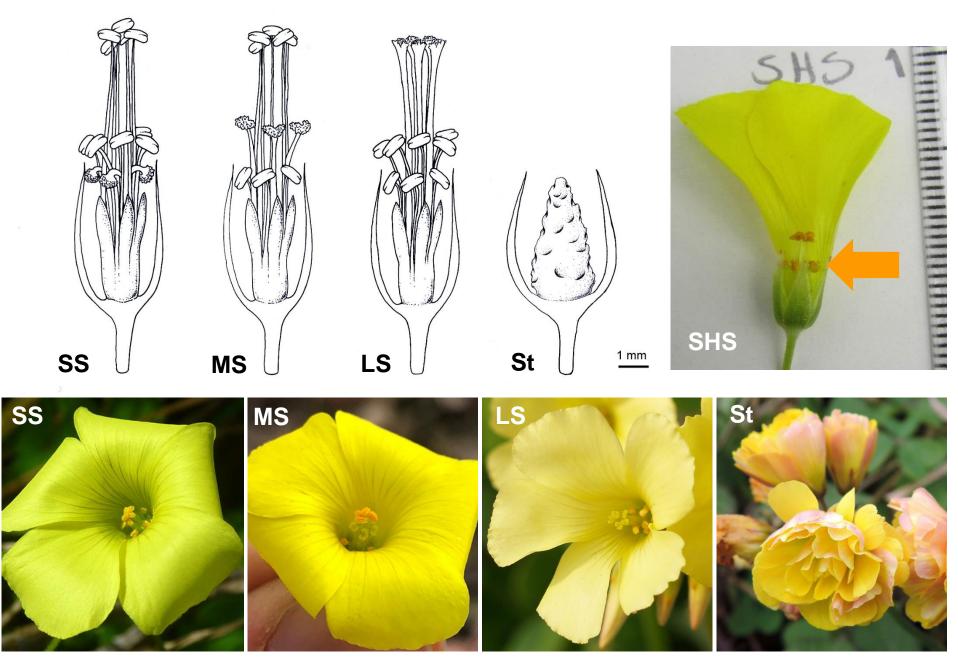
Participants will be included as co-authors in the manuscript describing current patterns in the Mediterranean basin

Additionally, participants can be included in further works upon their involvedness in the project





# Looking forward for your participation!



**Figure 1.** Bermuda buttercup (*Oxalis pes-caprae*) floral forms: short- (SS), mid- (MS) and long-styled (LS) morphs, multipedal sterile form (St) and semi-homostylous morph (SHS).